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DNA Binding and Antimicrobial Studies on Co (III) and Fe (II) Metal Complexes Containing Mixed Ligands

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ABSTRACT

The DNA binding ability of the novel complexes $\text{Co}(\text{L}_1)_2\text{L}_2(\text{PF}_6)_3$ [Complex (1)] and $\text{Fe}(\text{L}_1)_2\text{L}_2(\text{PF}_6)_2$ [Complex (2)] containing bioactive mixed ligand of type $\text{L}_1=8$ -Hydroxyquinoline and $\text{L}_2=1,10$ -Phenanthroline have been synthesized and characterized by analytical and spectral methods. The intrinsic binding constant K_b has been estimated at room temperature. The binding constant of the complexes in 5 mM Tris-HCl/50 mM NaCl buffer at pH 7.2, are $26.64 \times 10^6 \text{ M}^{-1}$ and $23.65 \times 10^6 \text{ M}^{-1}$ for complex (1) and (2) respectively. The absorption spectra indicate that the complexes intercalate between the base pairs of the CT-DNA tightly. The synthesized compounds have been tested against microorganisms such as Gram positive bacteria (*Staphylococcus aureus*), Gram-negative bacteria (*Escherichia coli*, *Proteus vulgaris* and *Pseudomonas aeruginosa*) and fungus (*Aspergillus niger*). A comparative study of the Minimum Inhibitory Concentration (MIC) values of the ligands and their complexes indicates that the complexes exhibit moderate antimicrobial activity than the free ligand and control.

Keywords: Co(III) & Fe(II) complexes, Ligands, DNA Binding, Antimicrobial studies.

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INTRODUCTION

Organometallic chemistry has been developed, in the last four decades, to be the largest and important branch as a link connecting the fields of organic and inorganic chemistry. One of the major applications of the transition metal complexes is its medical testing as antibacterial and antitumor agents aiming toward the discovery of an effective and safe therapeutic regimen for the treatment of bacterial infections and cancers. In addition, a great many Schiff' base complexes with metals have also provoked wide interest because they possess a diverse spectrum of biological and pharmaceutical activities, including antitumor and antioxidative activities. Moreover, mixed-ligand complexes are observed in biological systems or in the intermediate chemical reactions with metal ions, which are important to understand the respective chemistry. Investigations concerning 1,10- phenanthroline mixed-ligand chelate systems would provide toward understanding the driving forces that led to the formation of such mixed ligand complexes [1]. Moreover, Cu(II) and Zn(II) complexes with oxygen, sulphur and nitrogen containing ligands are the subject of intensive biological evaluation in the search for less toxic and more selective anticancer therapies [2-3].

Schiff bases are considered as a very important class of organic compounds which have wide applications in many biological aspects. Some Schiff bases were reported to possess antibacterial, antifungal and antitumor activities [4]. Due to their multiple implications, the transition metal complexes with Schiff bases, as ligands, are of paramount scientific interest. Schiff bases with donors (N, O) have structure similarities with natural biological systems and due to the presence of imine group (the $-N=CH-$), are utilized in elucidating the mechanism of transformation and rasemination reaction in biological systems [5]. Schiff base complexes have been used as drugs. Moreover, it is well known that some drug activities, when administered as metal complexes, are being increased, and several Schiff base complexes have also been shown to inhibit tumor growth. The effect of the presence of methyl substituent in the phenyl rings of aromatic Schiff bases on their antimicrobial activity has been reported [6]. Moreover, the incorporation of transition metal into Schiff bases enhances the biological activity of the ligand and decreases the cytotoxic effects of both the metal ion and ligand on the host [7].

Recently a novel Schiff base ligand has been synthesized and characterized by Raman et al. But, the antimicrobial activity for these new mixed ligand complexes of the above Schiff base has not been done. Hence, in this work, its complexes have been synthesized and the in vitro antimicrobial activity of all the above compounds has been carried out against three bacteria, and two fungi [8-9].

The study of mixed ligand-complex formation is relevant in the field of analytical chemistry, where the use of mixed ligand complexes allows the development of methods with increased selectivity, sensitivity and has also great importance in the field of biological and environmental chemistry [10-11].

However, the literature survey that the mixed ligands of 8-Hydroxyquinoline and 1-10 Phenanthroline derivatives with their transition metal complexes have not been reported and

studied so far. Hence the present study aims for the Synthesis, Characterization, Antimicrobial and DNA binding studies of Co and Fe complexes containing mixed ligands 8-hydroxyquinoline and 1,10-Phenanthroline.

MATERIALS AND METHODS

Experimental

All reagents and solvents were of AR grade, solvents were purified and used. Metal salts such as Cobalt chloride and ferrous sulphate were purchased from Karnataka fine chemicals. Ligands such as 8-Hydroxyquinoline and 1, 10-Phenanthroline was purchased from Himedia chemicals Ltd. (Bangalore).

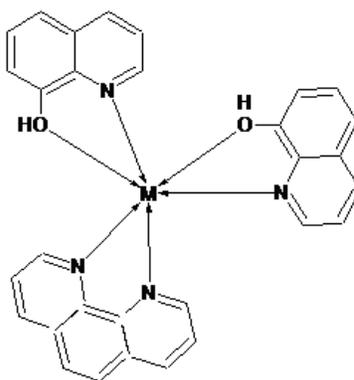
Synthesis of Metal Complexes

Synthesis of Co complex $[Co(L_1)_2L_2](PF_6)_3$ [Complex (1)]

Cobalt chloride (0.237g, 1mmol) 8-hydroxyquinoline L_1 (0.29g, 2mmol) 1,10-phenanthroline L_2 (0.234g, 1mmol) were dissolved in ethanolic solution and refluxed on the water bath for 4 hours. The contents were cooled to obtain precipitate. The complex was filtered and dried under vacuum before being recrystallized in acetone.

Synthesis of Fe complex $[Fe(L_1)_2L_2](PF_6)_2$ [Complex (2)]

Ferrous sulphate (0.278 g, 1mmol), 8-hydroxyquinoline L_1 (0.29 g, 2mmol), 1,10-phenanthroline L_2 (0.234g, 1mmol) were dissolved in ethanolic solution and refluxed on the water bath for 4 hours. The contents were cooled to obtain precipitate. The complex was filtered and dried under vacuum before being recrystallized in acetone.



Where M=Co or Fe

FIGURE 1: Proposed structure of the metal complexes

Spectral Measurements

IR spectra were recorded with Shimadzu model FT-IR spectrophotometer by using KBr pellets. UV-visible absorption spectra were recorded using ELICO model SL-159 UV-Vis Spectrophotometer at room temperature.

DNA Binding Studies

The concentration of CT-DNA per nucleotide $[C(p)]$ was measured by using its known extinction coefficient at 260 nm ($6600 \text{ M}^{-1} \text{ cm}^{-1}$)[10]. TrisHCl-buffer [5mM Tris(hydroxymethyl) amino methane, pH 7.2, 50 mM NaCl] was used for the absorption experiments.

Absorption titration experiments were carried out by varying the DNA concentration (0-100 μM) and maintaining the metal complex concentration constant. Absorption spectra were recorded after each successive addition of DNA and equilibration (approximately 10 minutes). For both (1) and (2), the observed data were then fitted into Equation (1) to obtain the intrinsic binding constant K_b [12].

$$[\text{DNA}]/(\epsilon_a - \epsilon_f) = [\text{DNA}]/(\epsilon_b - \epsilon_f) + 1/K_b (\epsilon_a - \epsilon_f),$$

Where ϵ_a , ϵ_b , and ϵ_f are the apparent, bound, and free metal complex extinction coefficients, respectively, at 322 nm for (1) and 328 nm for (2). A plot of $[\text{DNA}]/(\epsilon_b - \epsilon_f)$ versus $[\text{DNA}]$ gave a slope of $1/(\epsilon_b - \epsilon_f)$ and a y intercept equal to $1/K_b (\epsilon_b - \epsilon_f)$, where K_b is ratio of the slope to the y intercept.

Antimicrobial Studies

The ligands and its complexes were investigated for anti-bacterial and anti-fungal properties. Gram positive bacteria (*Staphylococcus aureus*), Gram-negative bacteria (*Escherichia coli*, *Proteus vulgaris* and *Pseudomonas aeruginosa*) and one fungus (*Aspergillus niger*) were used in this study to assess their antimicrobial properties. The tested compounds were dissolved in DMSO and the solutions were serially diluted in order to find the MIC values. The antibiotic Chloramphenicol was used as standard reference in the case of Gram-negative bacteria, Amikacin was used as standard reference in case of Gram-positive bacteria and Clotrimazole was used as standard anti-fungal reference. The solvent DMSO was used as negative control. In a typical procedure, a well was made in the Muller Hinton agar medium inoculated with microorganisms. The well was filled with the test solution using a micropipette and the plates were incubated at 37 °C at 72 h for fungi and 24 h for bacteria. During this period, the test solution diffused and affected the growth of the inoculated fungi and bacteria. Activity was determined by measuring the diameter of the zone of inhibition (mm).

RESULTS AND DISCUSSION

Characterization of Complexes

The elemental analysis data are agreed with the theoretical values within the limit of experimental error. These complexes are soluble in DMF, DMSO and in buffer (pH 7.2) solution.

The IR spectra of complexes were recorded in the range of 4000-400 cm^{-1} on KBr pellets. The spectra of the ligands 8-Hydroxyquinoline and 1,10-Phenanthroline showed bands at 1577 cm^{-1} and 1597 cm^{-1} assigned to $\nu\text{C}=\text{N}$ aromatic hydrocarbon, 3045 cm^{-1} and 3024 cm^{-1} assigned to $\nu\text{C}-\text{H}$ group and 3600 cm^{-1} and 3800 cm^{-1} assigned to $\nu\text{O}-\text{H}$ group. In the spectra of complexes **(1)** and **(2)**, these bands were shifted to 1550 cm^{-1} and 1499 cm^{-1} for $n(\text{C}=\text{N})$ group respectively. In addition, the IR spectrum of the PF6 salts of each complex showed a strong band at 727 cm^{-1} and 755 cm^{-1} ascribed to the counter anion and this band was absent for the corresponding salts [13].

Absorption Spectral Studies

The absorption spectra of complexes **(1)** and **(2)** in the absence and presence of CT-DNA are given in Figures 2 and 3, respectively. Figure 2 depicts a well-resolved band at 274 nm for complex **(1)** and in Figure 3; there exists a well-resolved band at 269 nm for complex **(2)** with increasing DNA concentrations (0–100 μM). The result shows that the absorbance (hypochromism) decreased by the successive addition of CT-DNA to the complex solution. The hypochromism observed for the bands of complexes **(1)** and **(2)** is accompanied by a small bathochromic shift of 2 and 1 nm, as shown in Figures 2 and 3, respectively. The hypochromism and bathochromic shift observed for the complexes suggest that binding is in intercalative mode. To compare quantitatively the DNA binding strengths of these complexes, the intrinsic DNA binding constants K_b are determined from the decay of the absorbance at 274 nm for complex **(1)** and 269 nm for complex **(2)** with increasing concentrations of DNA by using equation (1) given in our previous article [14-19]. The observed K_b values for complexes **(1)** and **(2)** are equal to the classical intercalators bound to CT-DNA. The K_b values for complexes **(1)** and **(2)** are $26.64 \times 10^6 \text{ M}^{-1}$ and $23.65 \times 10^6 \text{ M}^{-1}$, respectively. Thus, it is obvious that the present complexes are involved in intercalative interactions with CT-DNA.

Antimicrobial Studies

The in vitro antimicrobial activity of the compounds was tested against the bacterial species including *Staphylococcus aureus*, *Proteus vulgaris*, *Escherichia coli*, *Pseudomonas aeruginosa* and the fungus *Aspergillus niger* by well diffusion method. These complexes are inhibiting Gram-positive and Gram negative bacterial strains. The importance of this unique property of the investigated Schiff base complexes lies in the fact that, it can be applied safely in the treatment of infections and some common diseases e.g. Septicaemia, Gastroenteritis, Urinary tract infections and hospital acquired infections [20]. The ligand and their complexes have been tested for in vitro growth inhibitory activity against gram-positive microbe

Staphylococcus aureus, and gram-negative microbe's *Escherichia coli*, *Proteus vulgaris*, *Pseudomonas aeruginosa* by using well-diffusion method. As the test solution concentration increases, the biological activity also increases. The Minimum Inhibitory Concentration (MIC) values of the investigated compounds are summarized in Table 1. From the table, the observed MIC values indicate that the Fe complex have higher antibacterial activity than Co complex. But the complexes showed no inhibition against fungi. (*Aspergillus niger*).

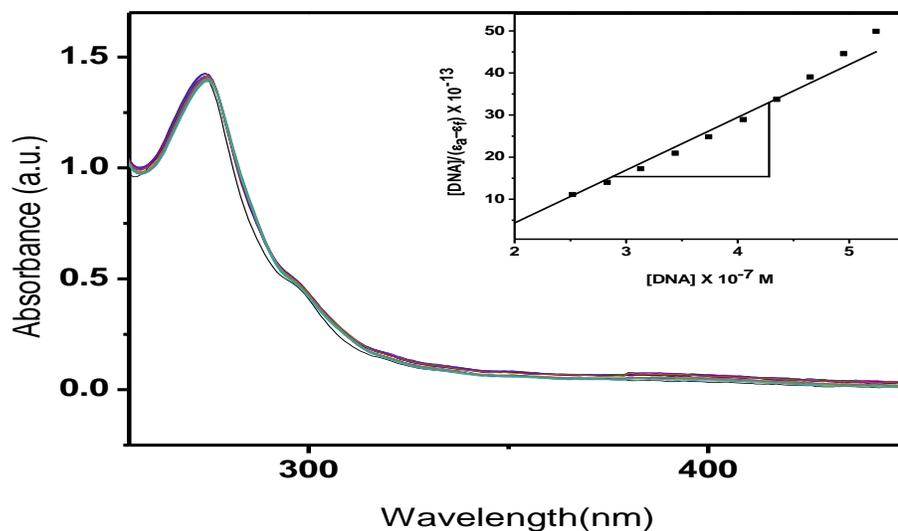


FIGURE 2: Absorption spectra of complex (1) in Tris-HCl buffer upon addition of DNA. $[Co] = 0.5 \mu M$, $[DNA] = 0-100 \mu M$. Arrow shows the absorbance changing upon the increase of DNA concentration. Inset: The plot of $[DNA]/(\epsilon_a - \epsilon_f)$ versus $[DNA]$ for the titration of DNA with Co(II) complex.

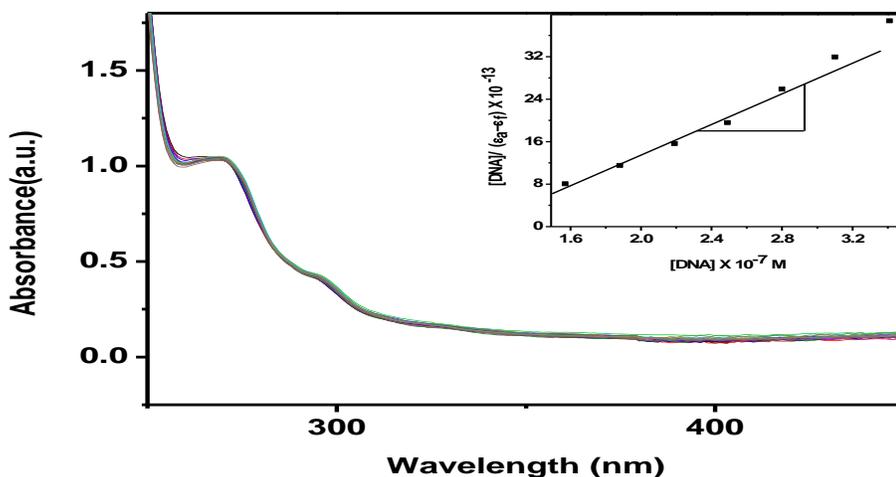


FIGURE 3: Absorption spectra of complex (2) in Tris-HCl buffer upon addition of DNA. $[Fe] = 0.5 \mu M$, $[DNA] = 0-100 \mu M$. Arrow shows the absorbance changing upon the increase of DNA concentration. Inset: The plot of $[DNA]/(\epsilon_a - \epsilon_f)$ versus $[DNA]$ for the titration of DNA with Fe(II) complex.

Table 1: Antimicrobial activity of the ligand and metal complexes

Sl.No	Extract	Conc (mg/ml)	<i>E.coli</i> (mm)	<i>P. aeruginosa</i> (mm)	<i>Staphylococcus aureus</i> (mm)	<i>Proteus vulgaris</i> (mm)	<i>A.niger</i> (mm)
1	L ₁	0.5	0	8	0	0	0
		1	0	15	0	0	0
		1.5	0	16	0	0	0
		2	0	17	12	0	0
		2.5	0	19	15	0	0
2	L ₂	0.5	0	0	0	12	0
		1	0	10	10	15	0
		1.5	15	12	16	16	0
		2	22	15	19	21	0
		2.5	24	22	23	22	0
3	Co(L ₁) ₂ L ₂	0.5	0	0	0	0	0
		1	0	0	0	0	0
		1.5	0	0	0	0	0
		2	0	0	0	0	0
		2.5	0	0	0	0	0
4	Fe(L ₁) ₂ L ₂	0.5	0	0	0	0	0
		1	0	10	0	0	0
		1.5	0	11	0	0	0
		2	0	15	16	0	0
		2.5	0	17	18	0	0
5	Antibiotic	0.5	21	24	21	21	0
		1	21	22	20	22	0
		1.5	22	22	21	22	0
		2	24	24	24	24	0
		2.5	22	24	25	24	0
6	DMSO	0.5	0	0	0	0	0
		1	0	0	0	0	0
		1.5	0	0	0	0	0
		2	0	0	0	0	0
		2.5	0	0	0	0	0

Less than 10mm-- Inactive; Less than 10-15mm--Weakly active; Less than 15-20mm-- Moderately active; More than 20mm—Highly active

Standard: Antibiotic Chloramphenicol for Gram negative bacteria, Amikacin for Gram-positive bacteria, Clotrimazole for fungi

Control: DMSO



CONCLUSION

In conclusion, we have synthesized and characterized two new complexes of the type $[\text{Co}(\text{L}_1)^2\text{L}_2] (\text{PF}_6)_3$ (**1**) and $[\text{Fe}(\text{L}_1)^2\text{L}_2] (\text{PF}_6)_2$ (**2**). Interactions of the new complexes with (double stranded) DNA were investigated by absorption spectra studies. From the experimental results, it was confirmed that the complexes bound with the double stranded DNA with binding constant $K_b = 26.64 \times 10^6 \text{ M}^{-1}$ and $23.65 \times 10^6 \text{ M}^{-1}$ (**2**), respectively. The complex (**1**) does not show any antibacterial and antifungal activity but complex (**2**) has antibacterial activity than the free ligands.

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REFERENCES

- [1] Raman N, Sobha S. J Serb Chem Soc 2010; 75: 1-16.
- [2] Jakupec MJ, Galanski M, Keppler BK. Rev Physiol Biochem Pharmacol 2003; 146:1–53.
- [3] a) Giovagnini L, Ronconi L, Aldinucci D, Lorenzon S, Sitran D, Fregona. J Med Chem 2005; 48: 1588–1595.
b) Gaballa AS, Asker MS, Barakat AS, Teleb SM. Spectrochim Acta 2007; 67A: 114-121.
- [4] Keskioglu E, Gunduzalp AB, Cete S, Hamurcu F, Erk B. Spectrochim Acta 2008; 70:634-640.
- [5] ShivaramaHolla B, Mahalinga M, SithambaramKarthikeyan M, Poojary B, Mohammed Akberalib P, Suchetha Kumar N. Eur J Med Chem 2005; 40:1173-1178.
- [6] Ameerunisha Begum MS, Saha S, Nethaji M, Chakravarty AR. J Inorg Biochem 2010; 104: 477-484.
- [7] Raman N, Selvan A, Pothiraj K, Jeyamurugan R. J Bloomers Res 2010; 3: 6-12.
- [8] Pyle AM, Barton JK, Lippard S J. J Am Chem Soc 1992; 114: 2303-2311.
- [9] Archika CB, Smita G, Anupa AK, Avinash SK, Vedavati G. Trans Metal Chem 2005; 30:312–316.
- [10] Tullius TD. American Chem Soc 2008; 402.
- [11] Reichmann ME, Rice SA, Thomas CA, Doty P. J Am Chem Soc 1954; 76: 3047–3053.
- [12] Wolfe A, Shimer GH, Meehan T. Biochemistry 1987; 26: 6392–6396.
- [13] Sreekanth B, Krishnamurthy G, BhojyaNaik HS, Vishnuvardhan TK, Vinaykumar B, Sharath N. Nucleosides Nucleotides Nucleic Acids 2011; 30: 83–96.
- [14] Sreekanth B, Krishnamurthy G, BhojyaNaik HS, Vishnuvardhan TK, Prabhakar MC. Synth React Inorg Met-Org Nano-Met Chem 2010; 40: 955–962.
- [15] Sreekanth B, Krishnamurthy G, Bhojya Naik HS, Vishnuvardhan TK, Lokesh MR, Shashikumar ND. Res J Pharm Biol and Chem Sci 2011; 2: 201-213.
- [16] Sreekanth B, Krishnamurthy G, BhojyaNaik HS, Vishnuvardhan TK, Lokesh MR, Shashikumar ND. J Chem and Pharm Res 2011; 3: 407-419.



- [17] Sreekanth B, Krishnamurthy G, Bhojya Naik HS, Vishnuvardhan TK. Nucleosides, Nucleotides and Nucliecacids 2012; 31: 1-13.
- [18] Sreekanth B, Krishnamurthy G, BhojyaNaik HS, Gopinath SM, Vishnuvardhan TK, Ismail Shareef M. Int J Pharma Biosci 2013; 4, 257-265.
- [19] VeenaPillai V, Sreekanth B. Int J of Pharma Biosci 2013; 4: 739-747.
- [20] Natesan M, Rajavel R. Int J of Pharma Tech 2010; 2: 1133-1157.